Neural Regulation of Sleep as Understood During Pre-Baldev Singh and Baldev Singh Eras

Velayudhan Mohan Kumar

Department of Physiology, All India Institute of Medical Sciences, New Delhi-110 029.

SUMMARY

Neurons in the midbrain reticular formation, primarily making up the ascending reticular activating system (ARAS), are important for production of low-voltage, fast-frequency, EEG pattern, commonly associated with wakefulness. The old notion that sleep is a passive phenomenon resulting from inactivation of the ARAS, is not true. There are specific brain regions that promote slow wave sleep (SWS) and others that are responsible for rapid eye movement (REM) sleep. Brain regions above the brain stem, namely the preoptic area (POA) of the hypothalamus, play an important role in the regulation of sleep. In spite of growing evidence in favour of the theory that the hypothalamus is the major centre regulating SWS, sleep-wake cycle, and even wakefulness, the recent findings confirm the important role of the brain stem in genesis of REM sleep. The importance which was previously given to the thalamus, for the regulation of SWS, is now seriously questioned, although it is still considered important for the genesis of the EEG spindles of normal sleep.

Chemical stimulation and neurotoxic lesion studies showed that the major function of the POA is sleep maintenance, rather than sleep initiation. Though there is interaction between thermoregulation and sleep regulation, there are different sets of neurons in the POA, endowed with the ability to regulate these two functions independently. At the same time, the POA could play an important role not only in integrating these two functions, but also in fine tuning the entire energy balance of the organism.

Key words: Sleep, Wakefulness, Preoptic area, Hypothalamus, REM sleep, Slow wave sleep, Brain stem, Thalamus, Ascending reticular activating system, EEG.

INTRODUCTION

This article is dedicated to the memory of Prof. Baldev Singh who had a deep interest in the field of sleep and consciousness. He had with him several books on sleep which were not available even in our libraries. Buying books was some kind of hobby for him. He used to lend those books to me, and discuss with me the theories of sleep prevalent during that period. Neurophysiology of sleep was a pet topic for him, and he used to select this subject, whenever he was invited to deliver lectures. The passion with which he spoke about this topic created such an interest in me that I took up this field of research and I have never deviated from it since then.

The understanding of neural regulation of sleep had undergone a drastic change in the last few decades. According to the classical concept of sleep which was prevalent till the sixties, which could be termed here as Pre-Baldev Singh Era, the neurons of the brain stem reticular formation played a major role in waking, consciousness and sleep. The later section of this article deals with the hypothalamic regulation of sleep, which dominated the scientific thinking during the Baldev Singh Era. The concept of sleep regulation by the brain stem, along with the latest modifications to the classical concepts, are also described here.

Brain stem regulation of wakefulness and passive deafferentation theory of sleep

Till around 1970, the sleep theory which dominated the text books were based on the findings of Moruzzi and Magoun (1). According to their findings, interruption of ascending flow of reticular impulses from from the brain stem (as it happens in the case of acute cerveau isole preparation), would result in a state of EEG recording which is similar to that of slow wave sleep (1). So, they concluded that the ascending influence, exerted by the brain stem on waking structures of

the cerebrum, maintains the wakefulness. The sudden withdrawal of the influence of the ascending reticular activating system (ARAS) would be the cause of slow wave EEG pattern obtained during sleep. It was then shown that the EEG pattern observed in acute cerveau isole (isolated brain) preparation is irreversible, similar to that found in a state of coma; rather than that of slow wave sleep (SWS). On the other hand, the EEG pattern observed in SWS is reversible. So, the original theory of withdrawal of ARAS influence was modified.

According to a new version of the old theory of passive deafferentation, a tonic ascending flow of reticular impulses would be responsible for the state of wakefulness, its complete interruption would lead to a state of coma. But, a temporary slackening of the tonic ARAS discharge might be responsible for the onset and the maintenance of natural sleep (2,3,4). These doctrines postulated that sleep is basically an inability to remain awake. When the tonic barrage falls below a critical level, the animal would fall asleep, simply because it is unable to stay awake.

Several years after being first proposed (1), the concept of a tonic reticular control of wakefulness and sleep still maintains some validity. Any level of cerebral activity is probably related to, and actually maintained by, a given level of reticular activation. ARAS projects to the thalamus and excite cells, which in turn send fibres to widespread areas of the cerebral cortex to produce the cerebral

cortical activation that occurs during wakefulness. In addition, brain stem reticular neurons project to the hypothalamus and other regions of the basal forebrain. The neurons which project from there to the cerebral cortex also participate in the maintenance of an "alert" state of the brain. In addition, there are also fibres projecting to the cortex, directly from the brain stem reticular formation. Changes in single unit and integrated discharge of the reticular system undoubtedly occur during the sleep-waking cycle (5,6,7). Whenever an arousal phenomenon interrupts the state of SWS, or when a fit of rage occurs against a background of relaxed wakefulness, there is phasic increase in the ARAS barrage. The level of reticular activity is assumed to be higher during wakefulness than during sleep (8,9,10). Both the mild tonic activation required for the maintenance of wakefulness, and the strong phasic discharge responsible for rage outbursts, require the support of the ARAS. The sudden withdrawal of the activating reticular influence would produce a striking imbalance, which might be responsible for the coma of the acute cerveau isole. The statement does not imply that the ARAS exerts a nonspecific facilitatory control upon the cerebrum (11).

Sleep generating mechanisms in the brain stem

Structures functionally antagonistic to the ARAS have been discovered in the lower brain stem and in the cerebrum itself. At least two of these regions are tonically active, as shown by the fact that their inactivation produces striking hyposomnia. They are localized in the lower brain stem and in the forebrain area (preoptic area). Several experiments suggest that deactivation is an active process, probably related to inhibition of the ascending reticular barrage. There is some electrophysiological evidence to show that the hypnogenic structures inhibit the ARAS (12,13). There are interrelations between activating and deactivating structures, and they have functional significance in sleep inducing mechanisms. Sleep could be broadly divided into two categories, i.e. slow wave sleep and rapid eye movement sleep.

Slow wave sleep (SWS): Several lines of evidence indicate that the nucleus of the solitary tract (NTS) is involved in sleep generation. Distention of the carotid sinus, a powerful stimulus for the NTS region, induced behavioural sleep. Low frequency stimulation of the vago-aortic nerve (which carry impulses from aortic sinus) produced slow waves in the EEG. Electrical stimulation of the NTS produced synchronization of the EEG and behavioural sleep, though high frequency stimulation of the same region produced arousal (14). Inactivation of the lower brain stem regions, including the NTS, produces a profound arousal.

There is some evidence that certain NTS neurons increase their discharge during SWS and that these neurons are reciprocally connected with the cells in the midbrain EEG arousal region. Thus,

Kumar et al (15) have localized the synchronizing structures at the nucleus gigantocellularis at the caudal brain stem. This study fulfilled the lacunae in our knowledge regarding the location of the regions in the caudal brain stem that bring about cortical EEG synchronization on electrical stimulation. It also characterized the features of synchronized waves elicited from those regions. Stimulation of ventromedial regions of the caudal brain stem, with low frequency, elicited stimulus-bound synchronized waves in the cortex, which were more prominent ipsilaterally. On the other hand, lowfrequency stimulation of dorsal and lateral areas produced synchronized waves which were either equally prominent on both sides, or more prominent on the contralateral side. The induced synchronized waves showed amplitude modulation and did not outlast the train of stimuli (15). These evidences indicate the strong possibility that these caudal brain stem regions play an important role in the genesis of sleep.

Rapid Eye Movement (REM) Sleep: The primary role for the genesis of SWS is played by structures above the brain stem, but the activity of certain groups of neurons in the brain stem is important in the generation of REM sleep. Some of these neurons are characterized as REMon neurons because of their selective activity during this state. Many of these neurons are located in the laterodorsal tegmental (LDT) and pedunculopontine tegmental (PPT) nuclei and use acetylcholine as a neurotransmitter. According to Datta (16), the cholinergic neurons involved in sleep-wakefulness are located at the peribrachial area (PBL). The major nuclei of the PBL are the PPT, the LTD, the cuneiform nuclei, and parts of the central tegmental field paralemniscal tegmental field. The activity of PBL cholinergic and noncholinergic cells are also responsible for the EEG activation process during wakefulness and REM sleep. During wakefulness, together with noradrenergic and serotonergic cells of the brain stem, PBL cholinergic and noncholinergic cells activate the diencephalic EEG desynchronizing structures, which in turn activate cortical neurons. However, during REM sleep, only PBL cholinergic and noncholinergic cells are responsible for activating those diencephalic EEG desynchronizing structures. An about a particular and appropriate the state of the sta

Several findings suggest that there are two populations of cholinergic neurons which have different influences on sleep-wakefulness regulation. A population of cholinergic neurons in the brainstem which are most active during paradoxical sleep (PS-on neuron) are inhibited by carbachol and excited by bicuculline. Another population of neurons, which are active both during waking and paradoxical sleep, are inhibited by carbachol and excited by noradrenaline

and histamine. Non-cholinergic PS-on neurons are excited by carbachol and inhibited by noradrenaline (17).

According to McCarley (18) the LDT and PPT neurons activate the neurons located in the pontine reticular formation (PRF) which are the effector neurons for REM sleep phenomena. They begin to depolarize even before the onset of the polysomnographic signs of REM sleep. These neurons begin to discharge (i.e. generate action potentials) as REM sleep is approached, and the high level of discharge is maintained throughout REM sleep (18). PRF neurons are important for the rapid eye movements saccades and the PGO waves, which are the cardinal signs of REM sleep. A group of dorsolateral PRF neurons controls the muscle atonia of REM sleep, and these neurons become active just before the onset of muscle atonia. Neurons in the bulbar reticular formation are also important for muscle atonia.

Neurons in the locus ceruleus (using norepinephrine) and neurons in the dorsal raphe (using serotonin) have an opposite time course of activity compared to the LDT and the PPT neurons. They become selectively inactive during REM. They do suppress REM sleep-promoting activity of the LDT and the PPT neurons. These neurons, with an opposite discharge time course, are called REM-off neurons. REM-off neurons are most active in the waking state. Their discharge declines during slow-wave sleep, and are virtually silent during REM sleep. They resume discharge near the end of the REM sleep episode.

This inverse pattern of activity to REM-off neurons has led to the hypothesis that these neurons may be REM-suppressive and interact with REM-on neurons in control of the REM sleep cycle. This concept is indirectly supported by production of REM sleep on cooling (i.e. inactivating) the nuclei where REM-off neurons are found. In vitro data from Luebke and co-workers (19) have provided direct support for the inhibition of cholinergic LDT neurons by serotonin.

Several classes of neurons are REM-off. Apart from norepinephrine-containing neurons in the locus ceruleus, and serotonin-containing neurons located in the raphe system, the midline collection of neurons that extends from the bulb to the midbrain, with serotonin-containing neurons in the more rostral regions, and histamine-containing neurons in the posterior hypothalamus, are REM-off. But transection studies indicate that the histaminergic neurons are not essential for REM sleep.

Role of thalamus in the genesis of sleep

There are three main systems in the brain which may be regarded as responsible for, or at least contributing to, the onset of sleep. They are localized respectively in the basal forebrain, in the lower brain stem, and in the midline nuclei of the thalamus.

According to Koella (20), the thalamamus is the head ganglion of sleep. Koella brings several interesting considerations in support of this hypothesis originally put forward by

Hess. Probably the most important among them is the striking similarity between natural sleep and the sleep induced by stimulation of the thalamus. Koella regards the hypnogenic regions of the basal forebrain, including the anterior hypothalamus, and of the lower brain stem, as subordinate sleep controlling structures. At the same time, he recalls that Hess (21) obtained a syndrome of generalized muscular relaxation, without typical sleep behaviour, from an area closely related to the basal forebrain. According to Steriade (22), brain stem, diencephalic and basal forebrain systems influence the functional modes of thalamic and cortical neurons during behavioural states of vigilance. The posterior hypothalamus and the preoptico-anterior hypothalamus (PO-AH) can influence the neurons of the midline thalamus. A majority of the influenced neurons of the midline thalamus showed increased firing on stimulation of the posterior hypothalamus. Although the number of neurons showing increased or decreased firing on the preoptico-anterior hypothalamic stimulation were nearly equal, stimulus bound increased firing in many neurons was followed by a prolonged decreased firing. It is likely that the hypothalamo-thalamic circuit constitutes a parallel pathway to the reticulo-thalamic circuit for alteration of the cortical EEG (23).

Stimulation experiments of Hess and electrophysiological investigations on thalamic driving of electrocortical EEG spindles (24,25) had focused the attention of sleep physiologists on the thalamus

for several years. Thalamocortical projection exhibits two distinct states of activity (a) synchronized rhythmic activity in the form of delta, spindle, and other slow waves during EEG-synchronized sleep and (b) tonic activity during waking and rapid-eye-movement sleep. Spindle waves are generated largely through a cyclical interaction between thalamocortical and thalamic reticular neurons involving both the intrinsic membrane properties of these cells and their anatomical interconnections (26).

The old literature on the effects of thalamic lesions was reviewed by Knott et al (27). A careful re-examination of these findings showed that the old experiments were contaminated by lesions of the hypothalamus and are therefore of little interest for the physiology of sleep. Naquet et al (28), who were able to follow six completely thalamectomised cats for up to 25 days, showed that the sleep-waking cycle was still present in these animals, although the EEG spindles were absent during behavioural sleep. Clearly sleep may occur even in the absence of the thalamus.

There is as yet no evidence of a marked hyposomnia produced by lesion of the midline thalamic nuclei(29). According to Moruzzi (29) the ARAS and a group of neurons lying in the posterior hypothalamus are probably concerned with the maintenance of wakefulness. The lower brain stem and the basal forebrain area contain structures with an opposing function, which exert a tonic deactivating influence and lead ultimately to sleep.

The role of hypothalamus in sleep-wakefulness

The ability to maintain a state of wakefulness do reappear in the chronic cerveau isole preparation. This shows that structures endowed with an activating influence are present in the cerebrum. They are mainly, though probably not exclusively, localized in the posterior hypothalamus. It is likely that these activating structures coincide with, or are at least strongly related to, the hypothalamic centre which is responsible for the outbursts of sham rage after decortication and for the defense-aggression behaviour of the normal animal.

Though it was known for years that there are centres or regions in the brain which could play an active role in the genesis of sleep, it was never given that much importance as to merit a place in the text books of neuroscience and physiology. In fact it was the statement of Moruzzi himself, in his review article which appeared in 1972, which played a major role in informing the world about the importance of the preoptic area (POA) in the genesis of sleep (29). Attempts have been made to locate within the POA the critical area for genesis of sleep. But the areas involved are not likely to restricted within this area. So, people have referred to this hypnogenic area as the basal forebrain or the anterior hypothalamus. But some others have referred to the area as the preoptic-anterior hypothalamus (PO-AH). In this manuscript, an attempt is made to refer to the areas using the terms mentioned by the authors

themselves. Information gathered about the role of the hypothalamus in sleepwakefulness is listed under different techniques employed in collecting the evidences.

Brain lesions and section studies: von Economo (30) was the first to draw the attention of physiologists to involvement of the rostral hypothalamus in sleep, on the basis of his observations in some cases of encephalitis. On postmortem examination of brain material collected from cases of encephalitis lethargica, he described two symptomatic patterns of the disease, associated with two different localizations of inflammatory lesions in the nervous system. In those cases in which somnolence was the distinguishing symptom, the lesions were regularly found in the posterior wall of the third ventricle, continuing caudally to the level of the occulomotor nucleus. In contrast to this, there were other cases in which insomnia was observed. Inflammation in these latter patients was associated with the rostral hypothalamus, the tuberal region and the adjacent portion of the striatum. From these observations, von Economo concluded that the rostral hypothalamic zone was a part of a "sleep regulating center" which, when appropriately excited, actively inhibited the thalamus and cerebral cortex and caused "brain sleep". Therefore, he concluded that the rostral hypothalamus is a "sleep center". In 1936, Ingram and co-workers (31) reported that in cats a lesion between mammillary bodies and the third nerves, which involved the caudal hypothalamus and the upper part

of the mesencephalic tegmentum, led to a state that resembled catalepsy. In 1939, Ranson (30) demonstrated in rhesus monkeys, that bilateral lesions in the lateral hypothalamus which extended up to the mammillary bodies, produced a lethargic syndrome. He reported that there was a lack of motor initiative and the symptoms were similar to those observed in catalepsy.

After von Economo's observations (30), several studies were carried out to elucidate the role played by the POA in the regulation of sleep. In 1946, Nauta (33), employing the knife cut lesion technique, showed that the rats became insomniac, restless and irritable after the lesion of the POA. They reacted vigorously even to minor stimuli. Those rats which survived up to 13 days did not show a return to what he called the "capacity of sleeping". He thought that the POA was an important region for sleeping and he termed this area as "sleep centre". Nagel and Satinoff (34) reported hyperactivity in rats after bilateral electrolytic lesion of the mPOA. McGinty and Sterman (35) reported that large bilateral POA lesions produced complete sleeplessness in cats. Smaller lesions resulted in significant reduction in SWS as well as in REM sleep. The severity of sleep suppression was found to be related to the size and localization of the lesions placed specifically within the POA (36). These lesions shortened the mean periodicity of the sleep awake cycle with a decrease in SWS and no alteration in REM sleep (36). Neurotoxic lesion studies provided convincing evidence of the

involvement of the POA neurons in the regulation of sleep. Lesions produced by neurotoxins such as kainic acid, which spares the fibres of passage in the medial and lateral POA, reduced both SWS and REM sleep (37,38). Suppression of sleep produced by the NMDA lesion of the medial preoptic area (mPOA) neurons showed the importance of this area in the regulation of sleep (39). Persistence of hyposomnia for three weeks after the lesion further showed that the deficits produced by the mPOA lesions were not compensated for. Insomnia was primarily due to a reduction in the duration of SWS episodes. There was a trend towards a reduction in the frequency and duration of REM sleep episodes. Recovery of sleep in the lesioned rats, after the fetal neural tissue transplantation, indicated the vital role of the mPOA in the regulation of sleep (40).

c-Fos expression studies: c-fos expression is strongly induced by both spontaneous and forced wakefulness in many brain regions. c-Fos expression was considerably increased in regions involved in the regulation of S-W, such as the locus coeruleus and the mPOA. With cfos antisense injection in the mPOA, it was demonstrated that c-fos expression in this region is causally involved in sleep regulation. c-Fos expression in other areas, such as the cerebral cortex and the hippocampus, are explained as related to the functional consequences of prolonged wakefulness and to the need for sleep (41). Periods of wakefulness result in the induction of immediate-early gene c-fos in the mPOA. Injections in the rat mPOA of c-fos antisense, oligonucleotides, blocked the expression of Fos protein detected immunocytochemically. Rats receiving bilateral antisense injections showed a higher percentage of wakefulness, the day after the injection, than controls receiving sense or sham injections or antisense injections outside the POA. These results suggest that blocking the expression of fos protein in the POA may interfere with the homeostatic regulation of sleep and wakefulness (42). Immunocytochemistry was used to identify the fos protein, an immediate-early gene product, in a group of ventrolateral POA neurons that is specifically activated during sleep. The retrograde tracer cholera toxin B, in combination with fos immunocytochemistry, was used to show that sleep-activated ventrolateral POA neurons innervate the tuberomammillary nucleus, a posterior hypothalamic cell group, thought to participate in the modulation of arousal. This monosynaptic pathway in the hypothalamus may play a key role in determining sleep-wake states (43).

Stimulation studies: Sterman and Clemente (44), on the basis of behavioural and electrophysiological observations, reported that bilateral stimulation of the POA in unanaesthetised freely moving cats, produced sleep. Low frequency stimulation was effective in inducing sleep. The effect of low frequency stimulation (5-25 cycles/sec) on induction of sleep was confirmed by Hernandez-Peon (45), and Yamaguchi et al (46). In addition, they showed that high frequency stimulation (200-300 cycles/sec) induced

cortical EEG desynchronization and some signs indicative of behavioural arousal. Thus, it can be concluded that both sleep and arousal responses can be obtained from electrical stimulation of the POA, depending upon the rate and site of stimulation.

Local intracerebral injection studies: Injection of norepinephrine (NE) at the mPOA produced arousal and the alpha receptors were involved in this response (47,48). Locally applied NE can act on both the presynaptic and postsynaptic receptors (49). It was later shown that the administration of NE did not induce arousal when the noradrenergic fibres in the mPOA were lesioned (49). These findings showed that the NE induces arousal by acting on the presynaptic alpha-2 receptors. Stimulation of alpha-2 receptors by externally applied NE and clonidine, would produce decreased release of endogenous NE from the presynaptic terminals (50). So, it is very likely that the decreased release of endogenous NE produced arousal in normal animals. This assumption was further supported by the finding that the alpha-2 agonist, clonidine, also produced arousal when applied at the mPOA (51). Alpha-2 receptors are primarily present on the presynaptic noradrenergic terminals (50,52). The presynaptic site of the action of clonidine was confirmed by the studies on VNA lesioned animals (53). yohimbine, an alpha-2 Similarly, antagonist, induced sleep by its action on the presynaptic terminals, as the effect was attenuated after the VNA lesion. Thus, the observations in the VNA

lesioned rats further confirmed the contention that these responses are mediated through presynaptic alpha-2 terminals (49,51).

Injection of beta adrenergic blocker, propranolol, at the mPOA was ineffective in changing sleep-wakefulness (47,54). But, beta receptors are also thought to be indirectly involved in the regulation of sleep-wakeful function. Beta agonist, isoproterenol injection into the mPOA induced wakefulness (55). Beta receptors in the mPOA have been shown to be important in inducing sexual arousal (56), which would in turn cause generalized arousal. So, it is possible that the injected isoproterenol would have produced arousal through a stimulatory action on the sexual arousal system. It may be hypothesized that there are different sets of neurons in the mPOA, controlling sleep, sex drive and other functions. It could be possible that the sleep regulating neurons are primarily stimulated through alpha receptors, whereas the neurons controlling the sex drive are activated through beta receptors.

Progesterone has also been reported to induce sleep, when applied directly into the rostral POA or basal forebrain area in cats, whereas no change was observed when it was injected into the caudal or lateral regions (57).

Prostaglandin has been shown to alter sleep by its action at the POA Prostaglandin D2 (PGD2) is present in high concentration in the hypothalamus in the rat brain. Ueno et al (58) reported that PGD2 application into the POA of

conscious rats increased SWS. On the other hand prostaglandin E2 (PGE2) produced arousal in rats (59,60). According to Hayaishi (61) PGD2 is the endogenous sleep inducing substance in rats, mice, monkeys and probably in humans. PGD synthase (PGDS), the enzyme that produces PGD2 in the brain, is the key enzyme in sleep regulation. When the enzyme activity is inhibited by its specific inhibitor, SeCl4 in vivo, rats can no longer sleep. PGDS is present mainly in the arachnoid membrane and choroid plexus. It is secreted into the cerebrospinal fluid to become beta-trace. PGD2 thus produced is bound to the receptors on the surface of the ventromedial region of the rostral basal forebrain. This signal is probably transmitted into the brain parenchyma by adenosine via adenosine A2a receptors. PGE2 plays a major role in the maintenance of wakefulness.

Hernandez-Peon (62) showed that the application of ACh at the POA elicited EEG synchronization and sleep, in cats, whereas carbachol, a cholinergic agonist, when applied at the mPOA, produced a fall in rectal temperature and an injection bound long lasting arousal in rats (63). Administration of GABA at the mPOA did not produce any significant alteration in S-W (64). Application of 5-HT crystals in the POA produced drowsiness and SWS in freely moving rats (46). But, Datta et al (65) showed that 5-HT application at the same site did not have any change in SW. Intracerebral injection of melatonin into the POA has been shown to increase SWS and REM sleep in rats (66).

Single unit recording studies: Mallick et al (67) showed that a majority (55%) of neurons of the POA showed alterations in their firing rate during transient changes in EEG. Among these 62.5% showed increased firing during synchronization and the remaining 37.5% showed increased firing during desynchronization of the EEG. Findlay and Hayward (68) showed that the majority of the neurons in the hypothalamus, including POA in freely moving rabbits, showed an increase in their firing rates during sleep as compared to the awake state. The POA neuronal activity was recorded during all states of sleep and wakefulness, and were classified into five groups according to their firing behaviour in relation to sleepwakefulness states (69). One third of the neurons showed no clear correlation with the sleep-waking states. Out of the 65 neurons which showed changes in activity with sleep-wakefulness states, 26 were most active during REM sleep. There were 16 neurons which were most inactive during REM sleep, some of which were inactive during SWS also. Those which were specifically active during SWS were 14 in number. Nine were less active during SWS than during wakefulness and REM sleep. About one third of neurons which showed increased discharge with SWS and REM sleep, began to increase their sleep-related activity in advance of the shift of sleep-wakefulness state recognized in EEG. These results suggest that the PO-AH areas are involved, at least in rats, in regulation of not only SWS but also REM sleep. sand the sand of

The role of the mPOA in the maintenance of sleep-wakefulness

There was persistent reduction in sleep (light SWS, deep SWS and REM sleep) and increase in the awake periods in rats after the mPOA neuronal lesion using NMDA (38). In cats, light SWS was not much affected after electrolytic and neurotoxic lesion (35,70). So, there was reduction in deep SWS and REM sleep in both rats and cats, after the mPOA lesion (39,70). The reduction in REM sleep suggests that the integrity of the POA is important for the regulation of this phase of sleep also (70,71). It is worth noting that the majority of the POA neurons show higher firing rates during REM sleep, than during SWS (69,72).

There was a significant decrease in the duration of SWS episodes after the mPOA lesion in rats (39). There was no significant increase in SWS frequency after the mPOA lesion. Thus, the initiation of sleep was not affected significantly by the mPOA lesion. This indicates that it is the maintenance of SWS rather than its initiation, which seems to have been mainly affected after the mPOA lesion. Lucas and Sterman(36) have also reported that there was a decrease in SWS bout duration, and an increase in the number of transitions of sleep stages after the basal forebrain lesions in cats.

There was a reduction in sleep pressure after the mPOA lesion, which shortened the sleep episode duration (39). This produced a reduction in the duration and frequency of deep SWS and REM sleep episodes. Normally, deep SWS occurred after the animal had spent some time in slow SWS. Similarly REM sleep appeared after the animals had spent some time in deep SWS. Thus, the reduction in sleep episode duration produced a reduction in frequency and duration of deeper stages of sleep, namely deep SWS and REM sleep. Unlike in human beings, in rats and cats, REM sleep forms a part of the deeper stage of sleep. So, the studies on NMDA lesioned rats indicate the possibility that the mPOA neurons are important for the maintenance and initiation of REM sleep.

Marginal increase in the frequency and duration of awake episodes, after the mPOA lesion (39), could be due to a release of the waking mechanism from the inhibitory influence of the POA (73).

Role of the mPOA in circadian and sleep-wake rhythms

Circadian rhythms are major features of the adaptation to our environment (74). In mammals, circadian rhythms are generated and regulated by a circadian timing system. This system consists of entertainment pathway, pace-maker, and pace-maker output to effector systems that are under circadian control. The primary entertainment pathway is the retinohypothalamic tract, which terminates on the circadian pace-maker, the suprachiasmatic nuclei of the hypothalamus. The output of the suprachiasmatic nuclei is principally to the other nuclei of the hypothalamus, the midline thalamus, and the basal forebrain. This provides a temporal organization to the sleep-wake cycle, in addition to many other physiological, endocrine and psychomotor functions (74).

In young adult human beings, the sleep-wake cycle coincides with the circadian cycle. But in cats and rats, and even in children, they do not exactly coincide. They are polycyclic and go through several cycles of sleep during the day and night. But all the same, they do show a circadian variation in the amount of sleep during day and night. The effect of brain lesion on sleep-wakeful cycle could be better studied in polycyclic animals, where the circadian cycle and the sleep-wakeful cycle could be viewed separately.

Many regions of the brain have the ability to produce arousal, or sleep. But it is necessary to locate the brain areas essential to generate the normal pattern of sleep-wake cycle. Neural regulation of sleep is highly incomplete without a proper regulation of sleep-wake cycle itself. Study of the sleep-wakeful cycle would also help in looking at the change in the duration and frequency of sleep episodes, as mentioned in earlier section. The sleep-wake cycle could be due to the slow accumulation and dissipation of chemical products within well defined groups of neurons in the brain (75). Rhythmicity is potentially present both in the cerebrum and in the brain stem, as shown by the experiments on the chronic cerveau isole and chronic decerebrate preparations (29). It is difficult to state where rhythmicity arises in the normal

animal, but several considerations suggest that the cerebrum; especially the hypothalamus, is the most likely location. According to one theory the sleep-waking cycle would normally arise within the cerebrum, and they are controlled by the ascending flow of brain stem impulses from the ARAS and the sleep inducing structures of the lower brain stem. But the important role of the hypothalamus for sleep-wakefulness change is particularly emphasized by Kawamura (76) on the basis of experimental data. According to him the mechanism of sleep-wakefulness change, produced by the forebrain, does not depend on the lower brain stem structures and ARAS. One week after rostral midbrain transection, the isolated forebrain showed sleep-wakefulness change, with circadian rhythm. In this preparation, after additional bilateral preoptic or posterior hypothalamic lesions, ECoG "insomnia" or "coma" pattern appeared, respectively. Hypothalamic sleep-wakefulness mech-anism usually receives strong influence from the suprachiasmatic nucleus, but it can produce its own ultradian "rhythms" (sleep-wakefulness change), though very irregular, even without this input.

The mPOA lesion in rats produced a proportionate reduction in sleep during the day and night, without any obvious change in the day-night sleep ratio (39). Absence of any persistent change in the night-day ratio of sleep would suggest that the mPOA has no role to play in the circadian distribution of sleep. Asala et al (77) have reported an uneven suppression of sleep after the radiofrequency

lesion of the mPOA. In those rats the reduction in the light period sleep was compensated for by the dark period sleep. It is difficult to assign any reason for the differences in the observations in the radiofrequency lesioned and the neurotoxic lesioned rats. The damage to the input from suprachiasmatic nucleus in the radiofrequency lesioned rats, could have been instrumental in disrupting the nightday distribution of sleep. Night-day distribution of food and water intake was also not significantly altered after the mPOA lesion. Though there was no alteration in night-day sleep-wakefulness, food and water intake, the rest-activity cycle was disturbed by the mPOA lesion.

Inter-relationship between hypothalamic thermoregulatory and sleep regulatory mechanisms

Roberts and Robinson (78) have suggested that the POA thermoreceptors may provide input to the POA sleepregulating mechanisms. Stimulation of central receptors by changing blood temperature is likely to be an important source of impulses driving the sleep inducing structures of basal forebrain (30). It was hypothesised that the SWS in mammals and birds is controlled by thermoregulatory mechanisms (79). Local warming of the POA produces sleep (78,79,80). PO-AH warming increases EEG delta frequency activity during SWS (80). So, it was suggested that the PO-AH thermoregulatory mechanisms participate in the regulation of the depth of SWS. According to Nakao et al (81) the SWS is controlled by thermoregulatory mech40

anisms of the PO-AH. Circadian and homeostatic thermoregulatory processes may be integrated in this brain area. Sleep could be induced by radio frequency diathermic warming of the POA in cats and opossum (78). Cooling the POA produces huddled posture (82). Low ambient temperature suppressed sleep and mild environmental warming enhanced sleep in normal rats (83,84). Studies have shown that SWS is facilitated when brain temperature exceeds a threshold level (79). This threshold is hypothesized to be determined by responses of the PO-AH thermosensitive neurons and to be regulated by both circadian and homeostatic processes. SWS-induced brain and body cooling would provide several adaptations including lower energy utilization, reduced cerebral metabolism, protection of the brain against the sustained high temperatures of wakefulness, facilitation of immune defense processes and regulation of the timing of behavioural activity relative to the circadian light-dark cycle. So, it was suggested that the mPOA, anterior hypothalamic and basal forebrain network integrates thermor-egulatory and hypnogenic controls and induces EEG and behavioral deactivation, through suppression of the ARAS.

It has been demonstrated that there are neurons in the mPOA involved in the regulation of sleep and body temperature (85,86,87). Thermosensitive neurons of the PO-AH have been implicated in the regulation of both body temperature and SWS (88). The activation of sleep-related warm-sensitive neurons (WSN) and the

deactivation of wake-related cold-sensitive neurons (CSN) may play a key role in the onset and regulation of SWS (89). During SWS, a majority of WSN of the PO-AH exhibit increased discharge compared to wakefulness. CSN exhibit reduced discharge in SWS, compared to wakefulness. WSN with increased discharge in SWS exhibited increased thermosensitivity during SWS compared to wakefulness. CSN with decreased discharge during SWS exhibited decreased thermosensitivity in SWS. In addition, 9 out of 47 neurons that were thermoinsensitive during wakefulness became warm-sensitive during SWS. Changes in PO-AH neuronal thermosensitivity could be a component of the mechanism for stabilization of state after state transition change, with circulture rayther, in .(88)

WSN did not exhibit a significant change in thermosensitivity during REM sleep compared with wakefulness and SWS (90). In contrast, CSN exhibited decreased mean thermosensitivity during REM sleep compared with wakefulness. CSN as a group did not retain significant thermosensitivity in REM sleep. These findings are consistent with evidence that thermoeffector responses to cooling are lost in REM sleep, whereas some responses to warming are preserved (90).

The mPOA lesions do increase the rectal temperature (70,71,91,92). Hyperthermia observed during the first week after the mPOA lesion was severe. This was followed by a constant mild hyperthermia during the subsequent weeks (39,92,93). On the other hand, there was

no variation in the magnitude of reduction in sleep throughout the postlesion period. Thus, there was no temporal correlation between the sleep and temperature changes after the mPOA lesion. Though hyperthermia is the commonly reported observation, hypothermia was also reported after the lesion of the POA (94,95).

Though possible the strong interrelationship between the regulations body temperature and wakefulness has been suggested on the basis of single unit and local warming studies (79), they are not supported by intracerebral injection studies. Neurotransmitters and their antagonists, injected at the mPOA, could not always produce simultaneous alterations in sleep and body temperature (48,63,96). Carbachol administration at the mPOA produced hypothermia and arousal. But the changes in these physiological parameters did not have a temporal correlation. The arousal response outlasted far beyond the changes in body temperature (63). Administration of 5-HT at the mPOA produced hyperthermia without any change in sleep-wakefulness (65). So, it is possible that the changes in sleep-wakefulness resulting from the mPOA stimulation and lesion are not dependent on the body temperature changes. Thus, it may be suggested that the mPOA controls sleep and temperature through independent, but overlapping, neuronal circuits. This conclusion is also supported by the observations of Krueger and Takahashi (97).

Effects of noradrenaline (NA) on the activity of sleep-related neurons in the POA and the neighboring basal forebrain were examined in the rat by Osaka and Matsumura (86). Sleep-active neurons were generally inhibited by NA and the alpha 2-agonist clonidine, whereas the alpha 1-agonist methoxamine and the beta-agonist isoproterenol had no effect on them. Thus, alpha 2-receptors mediated the NA-induced inhibition. Wakingactive neurons were excited by NA and methoxamine, whereas isopro-terenol and clonidine were without effect. Accordingly, alpha 1-receptors probably mediated the NA-induced excitation. State-indifferent neurons, and REM sleepactive neurons were mostly insensitive to NA. These results suggest that NA promotes wakefulness by inhibiting sleepactive neurons and by exciting wakingactive neurons (86).

The noradrenergic terminals, when activated, bring about sleep hypothermia. On the basis of local application of NA, it has been postulated that there are also two separate groups of afferent noradrenergic inputs, ending on the mPOA neurons. One of them, terminating on sleep inducing neurons, is tonically active during sleep. Those afferents which synapses on temperature regulatory neurons are suggested to be normally inactive (53). Clonidine administration at the mPOA produced arousal (51), but it was ineffective in producing any change in temperature (98). Clonidine (alpha-2 agonist) injection into the mPOA, in

normal rats, resulted in the activation of presynaptic alpha-2 receptors, on both the groups of noradrenergic afferents, but it brought about decreased release of endogenous NA on those neurons in which there was a tonic release. This decreased release of endogenous NA produced arousal in sleeping animals (51). Clonidine also acted on the inactive terminals which synapse on the temperature regulatory neurons. Since these fibres normally secrete very little NA, there was no change in the body temperature when this drug was applied. Yohimbine, an alpha-2 antagonist, blocks the presynaptic receptors and facilitates the release of endogenous NA. Postsynaptic action of the released NE on alpha-1 receptors, induces sleep in normal animals (51,99,100). Yohimbine failed to exert facilitated release of NA from those fibres which synapse on to the temperature regulatory neurons, since they are normally inactive. Hence, there was no change in the body temperature on application of this drug.

Changes in body weight, food and water intake on sleep regulation

Earlier reports have shown that the alteration in food intake can disrupt sleep (101). There are reports in the literature which indicate that the REM sleep deprivation or total sleep deprivation increases the food intake (102,103,104). But the decrease in SWS and PS, resulting from the mPOA lesion, did not produce

any increase in food intake and water intake (39). Food deprivation in birds and squirrels resulted in a lowering of the thermoregulatory set point during sleep along with increased SWS (105).

Though there was no significant persistent change in food intake, there was a reduction in the body weight of the rats after the mPOA lesion with NMDA, and electrolytic lesion of the POA (39,84). Higher locomotor activity and increased body temperature, after the mPOA lesion, would produce increased energy expenditure. This might have resulted in a decrease in the body weight because there was no concomitant compensatory addition in energy intake (food intake), in spite of the increase in locomotor activity, rectal temperature and awake period. Therefore, after the lesion, the animal did not recognize low energy reserves, and so it did not bother to conserve energy. Thus, it can be hypothesized that the mPOA lesioned animals had lost the mechanism for the fine tuning of food intake regulation in response to the alteration in body homeostasis. The functional integrity of the mPOA may be essential for the regulation of food intake, in response to alterations in the temperature, locomotor activity and S-W. It can also be argued that the POA would normally facilitate sleep, an energy-conserving state, when energy reserves are at a critical level.

REFERENCES WAS SEENES OF A SERVICE

- Moruzzi G and Magoun HW (1949).
 Brain stem reticular formation and activation of the EEG. Electroenceph Clin Neurophysiol 1: 455-473.
- Kleitman N (1963). Sleep and wakefulness, 2nd ed. Chicago: Chicago University Press.
- 3 Moruzzi G (1963). The Physiology of sleep. *Endeavour* 22: 31-36.
- 4. Moruzzi G (1964). Active processes in the brain stem during sleep. *Harvey Lect* 58: 233-297.
- Huttenlocher PR (1961). Evoked and spontaneous activity in single units of medial brain stem during natural sleep and waking. J Neurophysiol 1: 405-419.
- 6. Schlag JD and Balvin R (1963).

 Background activity in the cerebral cortex and reticular formation in relation with the electroencephalogram. Exp. Neurol 8: 203-219.
- Podvoll EM and Goodman SJ (1967).
 Averaged neural electrical activity and arousal, Science 155: 223-225.
- 8. Bremer F (1954). Contribution & petude des mecanismes physiologiques du maintien de pactivite vigile due cerveau. Interaction de la formation reticulee et de Pecorce cerbrale dans le processus du reveil Arch Int Physiol 62: 157-178.
- 9. Dell P, Bonvallet M and Hugelin A (1961). Mechanisms of reticular deactivation. In: The nature of sleep, Wolstenholme GEW and O'Connor CM (eds). p86-107, London:Churchill.
- 10. Moruzzi G (1969). Sleep and instinctive behavior *Arch Ital Biol* **107**: 175-216.

- 11. Moruzzi G (1958). The functional significance of the ascending reticular system. *Arch Ital Biol* **96**: 17-28.
- 12. Bremer F (1970). Inhibitions intrathalamiques recurrentielles et physiologie du sommeil. Electroenceph Clin Neurophysiol 28: 1-16.
- 13. Bremer F (1970). Preoptic hypnogenic focus and mesencephalic reticular formation. *Brain Res* 21: 132-134.
- Magnes J, Moruzzi G and Pompeiano O (1961). Synchronisation of the EEG produced by low frequency electrical stimulation of the region of the solitary tract. Arch Ital Biol 99: 33-67.
- Kumar VM, Chhina GS and Singh B (1985). Mapping of areas in the caudal brain stem which produce stimulus bound synchronization in cortical EEG. Exp Neurol 89: 295-304.
- Datta S (1995). Neuronal activity in the peribrachial area: relationship to behavioral state control. Neurosci Biobehav Rev 19: 67-84.
- 17. Koyama Y, Kayama Y and Sakai K (1998). Neural mechanisms for sleep regulation. *Nippon Rinsho* **56**: 318-326.
- 18. Mccarley RW (1995). Sleep, dreams and states of consciousness. *Neurosci Med* **29**: 537-553.
- Luebke JI, Greene RW, Semba K, Kamondi A, McCarley RW and Reiner PB (1992). Serotonin hyperpolarizes cholinergic low threshold burst neurons in the rat laterodorsal tegmental nucleus in vitro. Proc Natl Acad Sci USA 89: 743.
- KoellaWA (1967).Sleep. Its nature and physiological organisation. Springfield: Ch C Thomas. 199 pp.

- 21. Hess WR (1944). Das Schlafsyndrom als Folge dienzephaler Reizung. Helv Physiol Pharmacol Acta 2: 305-344.
- 22. Steriade M (1993). Central core modulation of spontaneous oscillations and sensory transmission in thalamocortical systems. *Curr Opin Neurobiol* 3: 619-625.
- 23. Aleem A, Kumar VM, Ahuja GK and Singh B (1986). Influence of preoptico-anterior and posterior hypothalamus on midline thalamic neurons. *Brain Res Bull* 16: 545-548.
- Purpura DP and Yahr MD (1966). The Thalamus. New York and London. Columbia University Press, 438pp.
- Purpura DP (1968). Role of synaptic inhibition in synchronization of thalamocortical activity. *Prog Brain Res* 22: 107-122.
- 26. McCormick DA and Bal T (1997). Sleep and arousal: thalamocortical mechanisms. *Annu Rev Neurosci* **20**: 185-215
- Knott JR, Ingram, WR and Chiles WD (1955). Effects of subcortical lesions on cortical electroencephalogram in cats Arch Neurol Psychiat (Chic) 73: 203-215.
- 28. Naquet R, Lanoir J, and Albe-Fessard D (1965). Alterations transitores on definitives de zones diencephaliques chez le chat. Leurs effects ur l'activite electrique corticale et le sommeil. In: Aspects anatomo-functionnels de la physiologie du sommeil. Jouvet M (ed), Paris: Centre national de la Recherche Scientifique, 107-131.
- 29. Moruzzi G (1972). The sleep-waking cycle. *Ergeb Physiol* **64**: 1-67.
- 30. von Economo C (1929). Schlabheorie. Ergeb Physiol 28: 312-39.

- 31. Ingram WR, Barris RW and Ranson SW (1936). Catalepsy. An experimental study. *Arch Neurol Psychiat* (Chic) 35: 1175-1197.
- 32. Ranson SW (1939). Somnolence caused by hypothalamic lesions in the monkey. *Arch Neurol Psychiat (Chic)* **41**: 1-23.
- 33. Nauta WJH (1946). Hypothalamic regulation of sleep in rats. An experimental study. *J Neurophysiol* 9: 285-316.
- 34. Nagel JA and Satinoff E (1980). Mild cold exposure increases survival in rats with medial preoptic lesion. *Science* 208: 301-303.
- 35. McGinty D and Sterman MB (1968). Sleep suppression after basal forebrain lesions in the cat. *Science* **160**: 1253-1255.
- 36. Lucus EA and Sterman MB (1975). Effect of forebrain lesion on a polycyclic sleepwake cycle and sleep-wake patterns in cat. *Exp Neurol* 46: 368-388.
- Szymusiak R, McGinty D (1986). Sleep suppression following kainic acidinduced lesions of the basal forebrain. Exp Neurol 94: 598-614.
- John J, Kumar VM, Gopinath G, Ramesh V and Mallick HN (1994). Changes in sleep-wakefulness after kainic acid lesion of the preoptic area in rats. *Jpn J Physiol* 44: 231-242.
- 39. John J and Kumar VM (1998). Effect of NMDA lesion of medial preoptic neurons on sleep and other functions. Sleep 21: 585-597.
- John J, Kumar VM and Gopinath G (1998). Recovery of sleep after fetal preoptic transplantation in the medial preoptic area lesioned rats. Sleep 21: 598-603.

- 41. Pompeiano M, Cirelli C, Arrighi P and Tononi G (1995). c-Fos expression during wakefulness and sleep. *Neurophysiol Clin* **25**: 329-341.
- 42. Cirelli C, Pompeiano M, Arrighi P and Tononi G (1995). Sleep-waking changes after c-fos antisense injections in the medial preoptic area. *Neuroreport* 27: 801-805.
- 43. Sherin JE, Shiromani PJ, McCarley RW and Saper CB (1996). Activation of ventrolateral preoptic neurons during sleep. *Science* **271**: 216-219.
- 44. Sterman MB and Clemente CD (1962). Forebrain inhibitory mechanisms: sleep patterns induced by basal forebrain stimulation in the behaving cat. *Exp Neurol* 6: 103-117.
- 45. Hernandez-peon R (1962). Sleep induced by localized electrical or chemical stimulation of the forebrain, Electroenceph Clin Neurophysiol 14: 423-424.
- 46. Yamaguchi N, Marczynski TJ and Ling GM (1963). The effects of electrical and chemical stimulation of the preoptic region and some non-specific thalamic nuclei in unrestrained, waking animals. Electroenceph Clin Neurophysiol 15: 154.
- 47. Kumar VM, Datta S, Chhina GS, Gandhi N and Singh B. (1984) Sleep-awake responses elicited from medial preoptic area on application of norepinephrine and phenoxybenzamine in free moving rats. *Brain Res* 322: 322-325.
- Datta S, Kumar VM, Chhina GS and Singh B (1988). Interrelationship of thermal and sleep-wakefulness changes elicited from the medial preoptic area in rats. Exp Neurol 100: 40-50.

- 49. Kumar VM (1993). Noradrenaline mechanism in the regulation of sleepwakefulness: A special role at the preoptic area. In: Kumar VM, Mallick HN and Nayar U (eds), Sleepwakefulness, Wiley Eastern, New Delhi 25-34.
- 50. Starke K (1987). Presynaptic o-autoreceptors. *Rev Physiol Biochem Pharmacol* **107**: 73-145.
- 51. Ramesh V, Kumar VM, John J and Mallick HN (1995). Medial preoptic alpha-2 adrenoceptors in the regulation of sleep-wakefulness. *Physiol Behav* 57: 171-175.
- 52. Langer SZ (1981). Presynaptic regulation of the release of catecholamines. *Pharmacol Rev* **32**: 337-362.
- 53. Ramesh V and Kumar VM (1998). The role of alpha-2 receptros in the medial preoptic area in the regulation of sleep-wakefulness and body temperature.

 Neuroscience 85: 807-818.
- 54. Kumar VM, Datta S, Chhina GS and Singh B (1986). Alpha adrenergic system in medial preoptic area involved in sleep- wakefulness in rats. *Brain Res Bull* 16: 463-468.
- 55. Sood S, Dhawan JK, Ramesh V, John J, Gopinath G. and Kumar VM (1997). Role of medial preoptic area beta adrenoceptors in the regulation of sleepwakefulness. *Pharmacol Biochem Behav* 57: 1-5.
- Mallick HN, Manchanda SK and Kumar VM (1996). Beta adrenergic modulation of male sex behavior elicited from the medial preoptic area in rats. Behav Brain Res 74: 181-187.
- 57. Heuser G, Ling GM, and Kluver M (1967). Sleep induction by progesterone

- in the preoptic area in cats. Electroenceph Clin Neurophysiol 22: 122-127.
- 58. Ueno R, Ishikawa Y, Nakayama T and Hayaishi O (1982). Prostaglandiń D2 induces sleep when microinjected into the preoptic area of conscious rats. *Biochem Biophy Res Commun* **109**: 576-582.
- 59. Matsumura H, Goh Y, Ueno R, Sakai T and Hayaishi O (1988). Awaking effect of PGE2 microinjected into the preoptic area of rats. *Brain Res* **444**: 265-272.
- 60. Matsumura H, Honda K, Choi WS, Inoue S, Sakai T and Hayaishi O 1989). Evidence that brain prostaglandin E2 is involved in physiological sleep-wake regulation in rats. *Proc Natl Acad Sci* 86: 5666-5669.
- 61. Hayaishi O (1998). Prostaglandins and sleep. *Nippon Rinsho* 56: 285-289.
- 62. Hernandez-peon R (1965). Die neuralen Grundlagen des Schlafes. *Arzneimittel-Forsch* **15**: 1099-1118.
- 63. Talwar A and Kumar VM (1994). Effect of carbachol injection in the medial preoptic area on sleep-wakefulness and body temperature in free moving rats. *Ind J Physiol Pharmacol* **38**: 11-16.
- 64. Chari DM, Ramesh V, John J and Kumar VM (1995). Effect of application of gamma amino butyric acid at the medial preoptic area on sleep-wakefulness. *Ind J Physiol Pharmacol* **39**: 199-201.
- 65. Datta S, Kumar VM, Chinna GS and Singh B (1987). Effect of application of serotonin in the medial preoptic area on body temperature and sleep-wakefulness. *Ind J Exp Biol* **25**: 681-685.
- 66. Holmes SW and Sugden D (1982). Effects of melatonin on sleep and

- neurochemistry in the rat. *Br J Pharmacol* **76**: 95-101.
- Mallick BN, Chhina GS, Sundaram KR, Singh B and Kumar VM (1983). Activity of preoptic neurons during synchronization and desynchronization. *Exp Neurol* 81: 586-597.
- 68. Findlay ALR and Hayward JN (1969). Spontaneous activity of single neurones in the hypothalamus of rabbits during sleep and waking *J Physiol (Lond)* **201**: 237-258.
- 69. Koyama Y and Hayaishi O (1994). Firing of neurons in the preoptic/anterior hypothalamic areas in rat: its possible involvement in slow wave sleep. Neurosci Res 19: 31-38.
- 70. Sallanon M, Denoyer M, Kitalama C, Alibert N, Gay N and Jouvet M (1989).

 Long lasting insomnia induced by preoptic neuron lesions and its transient reversal by muscimol injection into the posterior hypothalamus in the cat. Neuroscience 32: 669-683.
- 71. Szymusiak R, Danowski J and McGinty D (1991) Exposure to heat restores sleep in cats with preoptic/anterior hypothalamic cell loss. *Brain Res* **541**: 134-138
- 72. Detari L, Juhasz G, and Kukorelli T (1984). Firing properties of cat basal forebrain neurons during sleep-wakeful cycle. Electroencephalogr Clin Neurophysiol 58: 362-368.
- 73. Bremer F (1975). Existence of mutual tonic inhibitory interaction between the preoptic hypnogenic structure and the midbrain reticular formation. *Brain Res* 96: 71-75.
- 74. Moore RY (1997). Circadian rhythms: basic neurobiology and clinical applications. *Annu Rev Med* **48**: 253-266.

- 75. Jouvet M (1972). The role of monoamines and acetylcholine-containing neurons in the regulation of the sleepwaking cycle. Ergeb Physiol 64: 166-307.
- 76. Kawamura H (1998). Physiology of sleep-wakefulness rhythms. Nippon Rinsho 56: 277-284
- 77. Asala SA, Okano Y, Honda K and Inoue S (1990). Effects of medial preoptic area lesion on sleep and wakefulness in unrestrained rats. *Neurosci Lett* **114**: 300-304.
- 78. Roberts WW and Robinson TCL (1969). Relaxation and sleep induced by warming of preoptic region and anterior hypothalamus in cats. *Exp Neurol* 25: 284-294.
- McGinty D and Szymusiak R (1990).
 Keeping cool: a hypothesis about the mechanisms and functions of slow-wave sleep. Trends Neurosci 13: 480-487.
- 80. McGinty D, Szymusiak R and Thomson D (1994). Preoptic/ anterior hypothalamic warming increases EEG delta frequency activity within non-rapid eye movement sleep. *Brain Res* 667: 273-277.
- 81. Nakao M, McGinty D, Szymusiak R and Yamamoto M (1995). A thermoregulatory model of sleep control. *Jpn J Physiol* **45**: 291-309.
- 82. Freeman WJ and Davis DD (1959). Effect on cats of conductive hypothalamic cooling. *Am J Physiol* **197**: 145-148.
- 83. Szymusiak R, Satinoff E, Schallert T and Whishaw IQ (1980). Brief skin temperature changes toward thermoneutrality trigger REM sleep in rats. *Physiol Behav* 25: 305-311.

- 84. Szymusiak R and Satinoff E (1984). Amibient temperature-dependence of sleep disturbance produced by basal forebrain damage in rats. *Brain Res Bull* 12: 295-305.
- 85. Osaka T and Matsumura H (1994). Noradrenergic inputs to sleep-related neurons in the preoptic area from the locus coeruleus and the ventrolateral medulla in the rat. Neurosci Res 19: 39-50.
- 86. Osaka T and Matsumura H (1995).

 Noradrenaline inhibits preoptic sleepactive neurons through alpha-2 receptors
 in the rat. *Neurosci Res* 21: 323-330.
- 87. Schmid HA and Pierau FK (1993). Temperature sensitivity of neurons in slices of the rat PO/AH hypothalamic area: effect of calcium. *Am J Physiol* **264**: R440-448.
- 88. Alam MN, McGinty D and Szymusiak R (1996). Preoptic/anterior hypothalamic neurons: thermosensitivity in wakefulness and non rapid eye movement sleep. *Brain Res* 29: 76-82.
- 89. Alam MN, McGinty D and Szymusiak R (1995). Neuronal discharge of preoptic/anterior hypothalamic thermosensitive neurons: relation to NREM sleep. *Am J Physiol* **269**: R1240-R1249.
- 90. Alam MN, McGinty D and Szymusiak R (1995). Preoptic/anterior hypothalamic neurons: thermosensitivity in rapid eye movement sleep. *Am J Physiol* **269**: R1250-R1257.
- 91. Satinoff E, Liran J and Clapman R (1982). Aberrations of circadian body temperature rhythms in rats with medial preoptic lesions. *Am J Physiol* **242**: R352-R357.

- 92. Kumar VM and Khan NA (1998). Role of the preoptic neurons in thermoregulation in rats. *Arch Clin Exp Med* 7: 24-27.
- 93. Kumar VM, John J, Govindaraju V, Khan NA and Raghunathan P. (1996). Magnetic resonance imaging of NMDA induced lesion of the medial preoptic area and changes in sleep, temperature and sex behaviour. *Neurosci Res* 24: 207-214.
- 94. Verma S, Kumar VM, Gopinath G, Sharma R and Tandon PN (1989). Recovery of preoptic-anterior hypothalamic functions after transplantation. Restorative Neurol Neuro Sci 1: 77-81.
- 95. Squires RD and Jacobson FH (1968). Chronic deficits of temperature regulation produced in cats by preoptic lesion. *Am J Physiol* **214**: 549-560.
- 96. Osborne P, Onoe H and Watanabe Y (1994). GABAergic system inducing hyperthermia in the rat preoptic area: its independence of prostaglandin E2 system. *Brain Res* **661**: 237-242.
- 97. Krueger JM and Takahashi S (1998).
 Thermoregulation and sleep. Closely linked but separable. *Ann N Y Acad Sci*15: 281-286
- 98. Tsoucaris-Kupfer D and Schmitt H (1972). Hypothermic effect of alphasympathomimetic agents and their

- antagonism by adrenergic and cholinergic blocking drugs. *Neuropharmocology* **11**: 625-635.
- 99. Garcia-Ladona FJ, Claro E, Garcia A and Picatoste F (1993). Denervation hypersensitivity of histamine H1-receptors in rat brain cortex. *Neuroreport* 4: 691-694.
- 100. Kumar VM, Sharma R, Wadhwa S and Manchanda SK (1993). Sleep inducing function of noradrenergic fibres in the medial preoptic area. *Brain Res Bull* 32: 153-158.
- 101. Danguir J and Nicolaidis S (1979).

 Dependence of sleep in nutrients availability. *Physiol Behav* 22: 735-740.
- 102. Dement W, Henry P, Cohen H and Ferguson J (1967). Studies on the effect of REM deprivation in humans and in animals. Res Publ Ass nerv ment Dis 45: 456-468.
- 103. Siegel JM (1975). REM sleep predicts subsequent food intake. *Physiol Behav* 15: 399-403.
- 104. Bhanot JL, Chhina GS, Singh B, Sachdeva U and Kumar VM (1989).

 REM sleep deprivation and food intake.

 Ind J Physiol Pharmacol 33: 139-145.
- 105. Berger RJ and Phillips NH (1988).

 Comparative aspects of energy metabolism, body temperature and sleep. Acta Physiol Scand 574: 21-27.

Announcement

From this issue onwards, the contents of Annals along with full-length journal articles can be viewed free of cost at the official website of the Academy at http://NAMSIndia.com